

Methylation

In the chemical sciences, **methylation** denotes the addition of a methyl group on a substrate, or the substitution of an atom (or group) by a methyl group. Methylation is a form of alkylation, with a methyl group replacing a hydrogen atom. These terms are commonly used in chemistry, biochemistry, soil science, and the biological sciences.

In biological systems, methylation is catalyzed by enzymes; such methylation can be involved in modification of heavy metals, regulation of gene expression, regulation of protein function, and RNA processing. In vitro methylation of tissue samples is also one method for reducing certain histological staining artifacts. The counterpart of methylation is called demethylation.

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In biology

In biological systems, methylation is accomplished by enzymes. Methylation can modify heavy metals, regulate gene expression, RNA processing and protein function. It has been recognized as a key process underlying epigenetics. The Methylation cycle in medicine relates to the metabolism of various systems including DN and the production of glutathione. Faulty methylation cycle has been related to various abnormal conditions including Myalgic Encephalomyelitis (ME CFS) ^[1]

Methanogenesis

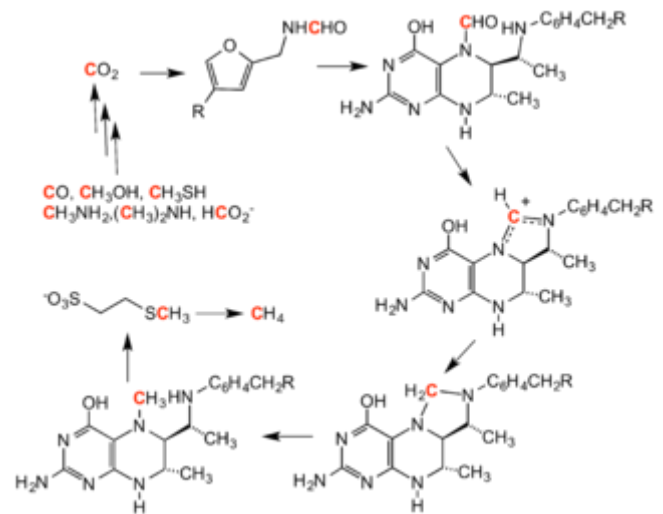
Methanogenesis, the process that generates methane from CO_2 , involves a series of methylation reactions. These reactions are effected by a set of enzymes harbored by a family of anaerobic microbes.^[2]

In reverse methanogenesis, methane serves as the methylating agent.

O-Methyltransferases

A wide variety of phenols undergo O-methylation to give anisole derivatives. This process, catalyzed by enzymes such as caffeoyl-CoA O-methyltransferase, is a key reaction in the biosynthesis of lignols, precursors to lignin, a major structural component of plants.

Plants produce flavonoids and isoflavones with methylations on hydroxyl groups, i.e. methoxy bonds. This 5-O-methylation affects the flavonoid's water solubility. Examples are 5-O-methylgenistein, 5-O-methylmyricetin or 5-O-methylquercetin, also known as azaleatin.



Cycle for methanogenesis, showing intermediates.

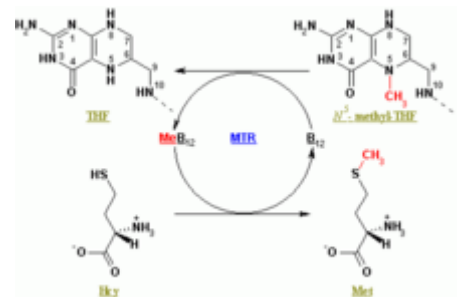
Proteins

Together with ubiquitin and phosphorylation, methylation is a major biochemical process for modifying protein function. The most prevalent protein methylations produce specific histones from arginine and lysine. Otherwise histidine, glutamate, asparagine, cysteine are susceptible to methylation. Some of these products include S-methylcysteine, two isomers of N-methylhistidine, and two isomers of N-methylarginine.^[3]

Methionine synthase

Methionine synthase regenerates methionine (Met) from homocysteine (Hcy). The overall reaction transforms 5-methyltetrahydrofolate ($\text{N}^5\text{-MeTHF}$) into tetrahydrofolate (THF) while transferring a methyl group to Hcy to form Met. Methionine Synthases can be cobalamin-dependent and cobalamin-independent: Plants have both, animals depend on the methylcobalamin-dependent form.

In methylcobalamin-dependent forms of the enzyme, the reaction proceeds by two steps in a ping-pong reaction. The enzyme is initially primed into a reactive state by the transfer of a methyl group from $\text{N}^5\text{-MeTHF}$ to Co(I) in enzyme-bound cobalamin (Cob), forming methylcobalamin (Me-Cob) that now contains Me-Co(III) and activating the enzyme. Then, a Hcy that has coordinated to an enzyme-bound zinc to form a reactive thiolate reacts with the Me-Cob. The activated methyl group is transferred from Me-Cob to the Hcy thiolate, which regenerates Co(I) in Cob, and Met is released from the enzyme.^[4]



The methylation reaction catalyzed by methionine synthase.

Heavy metals: arsenic, mercury, cadmium

Biomethylation is the pathway for converting some heavy elements into more mobile or more lethal derivatives that can enter the food chain. The biomethylation of arsenic compounds starts with the formation of methanearsonates. Thus, trivalent inorganic arsenic compounds are methylated to give methanearsonate. S-adenosylmethionine is the methyl donor. The methanearsonates are the precursors to dimethylarsonates, again by the cycle of reduction (to methylarsonous acid) followed by a second methylation.^[5] Related pathways apply to the biosynthesis of methylmercury.

Epigenetic methylation

DNA/RNA methylation

DNA methylation in vertebrates typically occurs at CpG sites (cytosine-phosphate-guanine sites—that is, where a cytosine is directly followed by a guanine in the DNA sequence). This methylation results in the conversion of the cytosine to 5-methylcytosine. The formation of Me-CpG is catalyzed by the enzyme DNA methyltransferase. In mammals, DNA methylation is common in body cells,^[6] and methylation of CpG sites seems to be the default.^{[7][8]} Human DNA has about 80–90% of CpG sites methylated, but there are certain areas, known as CpG islands, that are CG-rich (high cytosine and guanine content, made up of about 65% CG residues), wherein none is methylated. These are associated with the promoters of 56% of mammalian genes, including all ubiquitously expressed genes. One to two percent of the human genome are CpG clusters, and there is an inverse relationship between CpG methylation and transcriptional activity. Methylation contributing to epigenetic inheritance can occur through either DNA methylation or protein methylation. Improper methylations of human genes can lead to disease development,^{[9][10]} including cancer.^{[11][12]} Similarly, RNA methylation occurs in different RNA species viz. tRNA, rRNA, mRNA, tmRNA, snRNA, snoRNA, miRNA, and viral RNA. Different catalytic strategies are employed for RNA methylation by a variety of RNA-methyltransferases. RNA methylation is thought to have existed before DNA methylation in the early forms of life evolving on earth.^[13]

N6-methyladenosine (m6A) is the most common and abundant methylation modification in RNA molecules (mRNA) present in eukaryotes. 5-methylcytosine (5-mC) also commonly occurs in various RNA molecules. Recent data strongly suggest that m6A and 5-mC RNA methylation affects the regulation of various biological processes such as RNA stability and mRNA translation,^[14] and that abnormal RNA methylation contributes to etiology of human diseases.^[15]

Protein methylation

Protein methylation typically takes place on arginine or lysine amino acid residues in the protein sequence.^[16] Arginine can be methylated once (monomethylated arginine) or twice, with either both methyl groups on one terminal nitrogen (asymmetric dimethylarginine) or one on both nitrogens (symmetric dimethylarginine), by protein arginine methyltransferases (PRMTs). Lysine can be methylated once, twice, or three times by lysine methyltransferases. Protein methylation has been most studied in the histones. The transfer of methyl groups from S-adenosyl methionine to histones is catalyzed by enzymes known as histone methyltransferases. Histones that are methylated on certain residues can act epigenetically to repress or activate gene expression.^{[17][18]} Protein methylation is one type of post-translational modification.

Evolution

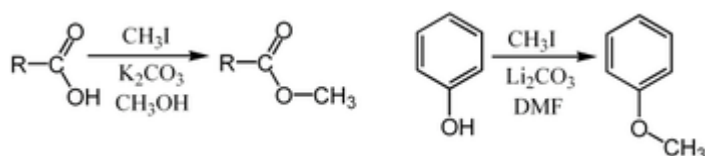
Methyl metabolism is very ancient and can be found in all organisms on earth, from bacteria to humans, indicating the importance of methyl metabolism for physiology.^[19] Indeed, pharmacological inhibition of global methylation in species ranging from human, mouse, fish, fly, round worm, plant, algae and cyanobacteria causes the same effects on their biological rhythms, demonstrating conserved physiological roles of methylation during evolution.^[20]

In chemistry

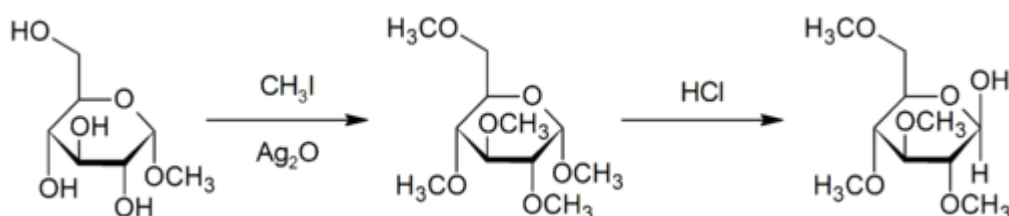
The term methylation in organic chemistry refers to the alkylation process used to describe the delivery of a CH₃ group.^[21]

Electrophilic methylation

Methylations are commonly performed using electrophilic methyl sources such as iodomethane,^[22] dimethyl sulfate,^{[23][24]} dimethyl carbonate,^[25] or tetramethylammonium chloride.^[26] Less common but more powerful (and more dangerous) methylating reagents include methyl triflate,^[27] diazomethane,^[28] and methyl fluorosulfonate (magic methyl). These reagents all react via S_N2 nucleophilic substitutions. For example, a carboxylate may be methylated on oxygen to give a methyl ester; an alkoxide salt RO⁻ may be likewise methylated to give an ether, ROCH₃; or a ketone enolate may be methylated on carbon to produce a new ketone.

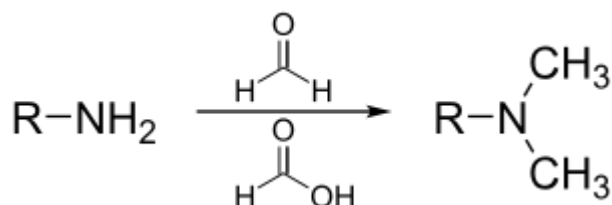


The Purdie methylation is a specific for the methylation at oxygen of carbohydrates using iodomethane and silver oxide.^[29]



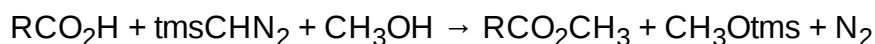
Eschweiler–Clarke methylation

The Eschweiler–Clarke reaction is a method for methylation of amines.^[30] This method avoids the risk of quaternization, which occurs when amines are methylated with methyl halides.



Diazomethane and trimethylsilyldiazomethane

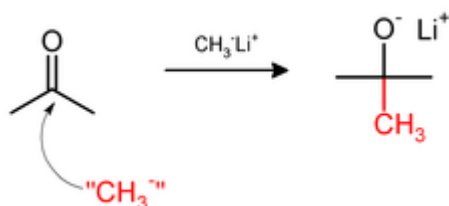
Diazomethane and the safer analogue trimethylsilyldiazomethane methylate carboxylic acids, phenols, and even alcohols:



The method offers the advantage that the side products are easily removed from the product mixture.^[31]

Nucleophilic methylation

Methylation sometimes involve use of *nucleophilic* methyl reagents. Strongly nucleophilic methylating agents include methyllithium (CH_3Li)^[32] or Grignard reagents such as methylmagnesium bromide (CH_3MgX).^[33] For example, CH_3Li will add methyl groups to the carbonyl ($\text{C}=\text{O}$) of ketones and aldehyde.:



Milder methylating agents include tetramethyltin, dimethylzinc, and trimethylaluminium.^[34]

See also

Biology topics

- Bisulfite sequencing – the biochemical method used to determine the presence or absence of methyl groups on a DNA sequence
- MethDB DNA Methylation Database
- Microscale thermophoresis – a biophysical method to determine the methylation state of DNA^[35]

Organic chemistry topics

- Alkylation
- Methoxy
- Titanium–zinc methylenation
- Petasis reagent
- Nysted reagent
- Wittig reaction
- Tebbe's reagent

References

1. Van Konyenberg R Phoenix Rising , 2014

2. Thauer, R. K., "Biochemistry of Methanogenesis: a Tribute to Marjory Stephenson", *Microbiology*, 1998, volume 144, pages 2377-2406.
3. Clarke, Steven G. (2018). "The ribosome: A hot spot for the identification of new types of protein methyltransferases" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6036201>). *Journal of Biological Chemistry*. **293** (27): 10438–10446. doi:10.1074/jbc.AW118.003235 (<https://doi.org/10.1074%2Fjbc.AW118.003235>). PMC 6036201 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6036201>). PMID 29743234 (<https://pubmed.ncbi.nlm.nih.gov/29743234>).
4. Matthews, R. G.; Smith, A. E.; Zhou, Z. S.; Taurog, R. E.; Bandarian, V.; Evans, J. C.; Ludwig, M. (2003). "Cobalamin-Dependent and Cobalamin-Independent Methionine Synthases: Are There Two Solutions to the Same Chemical Problem?". *Helvetica Chimica Acta*. **86** (12): 3939–3954. doi:10.1002/hlca.200390329 (<https://doi.org/10.1002%2Fhlca.200390329>).
5. Styblo, M.; Del Razo, L. M.; Vega, L.; Germolec, D. R.; LeCluyse, E. L.; Hamilton, G. A.; Reed, W.; Wang, C.; Cullen, W. R.; Thomas, D. J. (2000). "Comparative toxicity of trivalent and pentavalent inorganic and methylated arsenicals in rat and human cells". *Archives of Toxicology*. **74** (6): 289–299. doi:10.1007/s002040000134 (<https://doi.org/10.1007%2Fs002040000134>). PMID 11005674 (<https://pubmed.ncbi.nlm.nih.gov/11005674>). S2CID 1025140 (<https://api.semanticscholar.org/CorpusID:1025140>).
6. Tost J (2010). "DNA methylation: an introduction to the biology and the disease-associated changes of a promising biomarker". *Mol Biotechnol*. **44** (1): 71–81. doi:10.1007/s12033-009-9216-2 (<https://doi.org/10.1007%2Fs12033-009-9216-2>). PMID 19842073 (<https://pubmed.ncbi.nlm.nih.gov/19842073>). S2CID 20307488 (<https://api.semanticscholar.org/CorpusID:20307488>).
7. Lister R, Pelizzola M, Dowen RH, Hawkins RD, Hon G, Tonti-Filippini J, Nery JR, Lee L, Ye Z, Ngo QM, Edsall L, Antosiewicz-Bourget J, Stewart R, Ruotti V, Millar AH, Thomson JA, Ren B, Ecker JR (November 2009). "Human DNA methylomes at base resolution show widespread epigenomic differences" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2857523>). *Nature*. **462** (7271): 315–22. Bibcode:2009Natur.462..315L (<https://ui.adsabs.harvard.edu/abs/2009Natur.462..315L>). doi:10.1038/nature08514 (<https://doi.org/10.1038%2Fnature08514>). PMC 2857523 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2857523>). PMID 19829295 (<https://pubmed.ncbi.nlm.nih.gov/19829295>).
8. Stadler MB, Murr R, Burger L, Ivanek R, Lienert F, Schöler A, van Nimwegen E, Wirbelauer C, Oakeley EJ, Gaidatzis D, Tiwari VK, Schübeler D (December 2011). "DNA-binding factors shape the mouse methylome at distal regulatory regions" (<https://doi.org/10.1038/nature11086>). *Nature*. **480** (7378): 490–5. doi:10.1038/nature11086 (<https://doi.org/10.1038%2Fnature11086>). PMID 22170606 (<https://pubmed.ncbi.nlm.nih.gov/22170606>).
9. Rotondo JC, Selvatici R, Di Domenico M, Marci R, Vesce F, Tognon M, Martini F (September 2013). "Methylation loss at H19 imprinted gene correlates with methylenetetrahydrofolate reductase gene promoter hypermethylation in semen samples from infertile males" (<https://academic.oup.com/humrep/article/27/12/3632/651064>). *Epigenetics*. **8** (9): 990–7. doi:10.4161/epi.25798 (<https://doi.org/10.4161%2Fepi.25798>). PMC 3883776 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3883776>). PMID 23975186 (<https://pubmed.ncbi.nlm.nih.gov/23975186>).
10. Rotondo JC, Bosi S, Bazzan E, Di Domenico M, De Mattei M, Selvatici R, Patella A, Marci R, Tognon M, Martini F (December 2012). "Methylenetetrahydrofolate reductase gene promoter hypermethylation in semen samples of infertile couples correlates with recurrent spontaneous abortion" (<https://academic.oup.com/humrep/article/27/12/3632/651064>). *Human Reproduction*. **27** (12): 3632–8. doi:10.1093/humrep/des319 (<https://doi.org/10.1093%2Fhumrep%2Fdes319>). PMID 23010533 (<https://pubmed.ncbi.nlm.nih.gov/23010533>).

11. Rotondo JC, Borghi A, Selvatici R, Magri E, Bianchini E, Montinari E, Corazza M, Virgili A, Tognon M, Martini F (2016). "Hypermethylation-Induced Inactivation of the IRF6 Gene as a Possible Early Event in Progression of Vulvar Squamous Cell Carcinoma Associated With Lichen Sclerosus". *JAMA Dermatology*. **152** (8): 928–33. doi:10.1001/jamadermatol.2016.1336 (<https://doi.org/10.1001%2Fjamadermatol.2016.1336>). PMID 27223861 (<https://pubmed.ncbi.nlm.nih.gov/27223861>).
12. Rotondo JC, Borghi A, Selvatici R, Mazzoni E, Bononi I, Corazza M, Kussini J, Montinari E, Gafà R, Tognon M, Martini F (2018). "Association of Retinoic Acid Receptor β Gene With Onset and Progression of Lichen Sclerosus-Associated Vulvar Squamous Cell Carcinoma" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6128494>). *JAMA Dermatology*. **154** (7): 819–823. doi:10.1001/jamadermatol.2018.1373 (<https://doi.org/10.1001%2Fjamadermatol.2018.1373>). PMC 6128494 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6128494>). PMID 29898214 (<https://pubmed.ncbi.nlm.nih.gov/29898214>).
13. Rana, Ajay K.; Ankri, Serge (1 January 2016). "Reviving the RNA World: An Insight into the Appearance of RNA Methyltransferases" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4893491>). *Front Genet*. **7**: 99. doi:10.3389/fgene.2016.00099 (<https://doi.org/10.3389%2Ffgene.2016.00099>). PMC 4893491 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4893491>). PMID 27375676 (<https://pubmed.ncbi.nlm.nih.gov/27375676>).
14. Choi, Junhong; leong, Ka-Weng; Demirci, Hasan; Chen, Jin; Petrov, Alexey; Prabhakar, Arjun; O'Leary, Seán E.; Dominissini, Dan; Rechavi, Gideon (February 2016). "N6-methyladenosine in mRNA disrupts tRNA selection and translation-elongation dynamics" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4826618>). *Nature Structural & Molecular Biology*. **23** (2): 110–115. doi:10.1038/nsmb.3148 (<https://doi.org/10.1038%2Fnsmb.3148>). ISSN 1545-9993 (<https://www.worldcat.org/issn/1545-9993>). PMC 4826618 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4826618>). PMID 26751643 (<https://pubmed.ncbi.nlm.nih.gov/26751643>).
15. Stewart, Kendal (15 September 2017). "Methylation (MTHFR) Testing & Folate Deficiency" (<https://web.archive.org/web/20171012202147/https://www.genomixnutrition.com/methylation-testing-s/2.htm>). Archived from the original (<https://www.genomixnutrition.com/methylation-testing-s/2.htm>) on 12 October 2017. Retrieved 11 October 2017.
16. Walsh, Christopher (2006). "Chapter 5 – Protein Methylation" (<http://www.roberts-publishers.com/walsh/chapter5.pdf>) (PDF). *Posttranslational modification of proteins: expanding nature's inventory*. Roberts and Co. Publishers. ISBN 978-0-9747077-3-0.
17. Grewal, S. I.; Rice, J. C. (2004). "Regulation of heterochromatin by histone methylation and small RNAs" (<https://zenodo.org/record/1258832>). *Current Opinion in Cell Biology*. **16** (3): 230–238. doi:10.1016/j.ceb.2004.04.002 (<https://doi.org/10.1016%2Fj.ceb.2004.04.002>). PMID 15145346 (<https://pubmed.ncbi.nlm.nih.gov/15145346>).
18. Nakayama, J. -I.; Rice, J. C.; Strahl, B. D.; Allis, C. D.; Grewal, S. I. (2001). "Role of Histone H3 Lysine 9 Methylation in Epigenetic Control of Heterochromatin Assembly". *Science*. **292** (5514): 110–113. Bibcode:2001Sci...292..110N (<https://ui.adsabs.harvard.edu/abs/2001Sci...292..110N>). doi:10.1126/science.1060118 (<https://doi.org/10.1126%2Fscience.1060118>). PMID 11283354 (<https://pubmed.ncbi.nlm.nih.gov/11283354>). S2CID 16975534 (<https://api.semanticscholar.org/CorpusID:16975534>).
19. Kozbial, P.Z.; Mushegian, A.R. (2005). "Natural history of S-adenosylmethionine-binding proteins" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1282579>). *BMC Struct Biol*. **5** (19): 19. doi:10.1186/1472-6807-5-19 (<https://doi.org/10.1186%2F1472-6807-5-19>). PMC 1282579 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1282579>). PMID 16225687 (<https://pubmed.ncbi.nlm.nih.gov/16225687>).

20. Fustin, J.M.; Ye, S.; Rakers, C.; Kaneko, K.; Fukumoto, K.; Yamano, M.; Versteven, M.; Grünewald, E.; Cargill, S.J.; Tamai, T.K.; Xu, Y.; Jabbur, M.L.; Kojima, R.; Lamberti, M.L.; Yoshioka-Kobayashi, K.; Whitmore, D.; Tammam, S.; Howell, P.L.; Kageyama, R.; Matsuo, T.; Stanewsky, R.; Golombek, D.A.; Johnson, C.H.; Kakeya, H.; van Ooijen, G.; Okamura, H. (2020). "Methylation deficiency disrupts biological rhythms from bacteria to humans" (<https://doi.org/10.1038/s42003-020-0942-0>). *Communications Biology*. **3** (211). doi:10.1038/s42003-020-0942-0 (<https://doi.org/10.1038/s42003-020-0942-0>). PMID 32376902 (<https://pubmed.ncbi.nlm.nih.gov/32376902>).
21. March, Jerry; Smith, Michael W (2001). *March's advanced organic chemistry: reactions, mechanisms, and structure*. New York: Wiley. ISBN 978-0-471-58589-3.
22. Vyas, G. N.; Shah, N. M. (1951). "Quinacetophenone monomethyl ether". *Organic Syntheses*. **31**: 90. doi:10.15227/orgsyn.031.0090 (<https://doi.org/10.15227/orgsyn.031.0090>).
23. Hiers, G. S. (1929). "Anisole". *Organic Syntheses*. **9**: 12. doi:10.15227/orgsyn.009.0012 (<https://doi.org/10.15227/orgsyn.009.0012>).
24. Icke, Roland N.; Redemann, Ernst; Wisegarver, Burnett B.; Alles, Gordon A. (1949). "m-Methoxybenzaldehyde". *Organic Syntheses*. **29**: 63. doi:10.15227/orgsyn.029.0063 (<https://doi.org/10.15227/orgsyn.029.0063>).
25. Tundo, Pietro; Selva, Maurizio; Bomben, Andrea (1999). "Mono-C-methylation of arylacetone nitriles and methyl arylacetates by dimethyl carbonate: a general method for the synthesis of pure 2-arylpropionic acids. 2-Phenylpropionic acid". *Organic Syntheses*. **76**: 169. doi:10.15227/orgsyn.076.0169 (<https://doi.org/10.15227/orgsyn.076.0169>).
26. Nenad, Maroš; Polanc, Slovenko; Kočevar, Marijan (2008). "Microwave-assisted methylation of phenols with tetramethylammonium chloride in the presence of K₂CO₃ or Cs₂CO₃". *Tetrahedron*. **64** (51): 11618–11624. doi:10.1016/j.tet.2008.10.024 (<https://doi.org/10.1016/j.tet.2008.10.024>).
27. Poon, Kevin W. C.; Albiniak, Philip A.; Dudley, Gregory B. (2007). "Protection of alcohols using 2-benzyloxy-1-methylpyridinium trifluoromethanesulfonate: Methyl (R)-(-)-3-benzyloxy-2-methyl propanoate". *Organic Syntheses*. **84**: 295. doi:10.15227/orgsyn.084.0295 (<https://doi.org/10.15227/orgsyn.084.0295>).
28. Neeman, M.; Johnson, William S. (1961). "Cholestanyl methyl ether". *Organic Syntheses*. **41**: 9. doi:10.15227/orgsyn.041.0009 (<https://doi.org/10.15227/orgsyn.041.0009>).
29. Purdie, T.; Irvine, J. C. (1903). "C.?The alkylation of sugars" (<https://zenodo.org/record/2039403>). *Journal of the Chemical Society, Transactions*. **83**: 1021–1037. doi:10.1039/CT9038301021 (<https://doi.org/10.1039/CT9038301021>).
30. Icke, Roland N.; Wisegarver, Burnett B.; Alles, Gordon A. (1945). "β-Phenylethyldimethylamine". *Organic Syntheses*. **25**: 89. doi:10.15227/orgsyn.025.0089 (<https://doi.org/10.15227/orgsyn.025.0089>).
31. Shioiri, Takayuki; Aoyama, Toyohiko; Snowden, Timothy (2001). "Trimethylsilyldiazomethane". *Encyclopedia of Reagents for Organic Synthesis. e-EROS Encyclopedia of Reagents for Organic Synthesis*. doi:10.1002/047084289X.rt298.pub2 (<https://doi.org/10.1002/047084289X.rt298.pub2>). ISBN 978-0471936237.
32. Lipsky, Sharon D.; Hall, Stan S. (1976). "Aromatic Hydrocarbons from aromatic ketones and aldehydes: 1,1-Diphenylethane". *Organic Syntheses*. **55**: 7. doi:10.15227/orgsyn.055.0007 (<https://doi.org/10.15227/orgsyn.055.0007>).
33. Grummitt, Oliver; Becker, Ernest I. (1950). "trans-1-Phenyl-1,3-butadiene". *Organic Syntheses*. **30**: 75. doi:10.15227/orgsyn.030.0075 (<https://doi.org/10.15227/orgsyn.030.0075>).
34. Negishi, Ei-ichi; Matsushita, Hajime (1984). "Palladium-Catalyzed Synthesis of 1,4-Dienes by Allylation of Alkenylalane: α-Farnesene". *Organic Syntheses*. **62**: 31. doi:10.15227/orgsyn.062.0031 (<https://doi.org/10.15227/orgsyn.062.0031>).

35. Wienken CJ, Baaske P, Duhr S, Braun D (2011). "Thermophoretic melting curves quantify the conformation and stability of RNA and DNA" (<http://nar.oxfordjournals.org/content/early/2011/02/04/nar.gkr035.full?keytype=ref&ijkey=FsmS2lhrsZ8wdBr>). *Nucleic Acids Research*. **39** (8): e52. doi:10.1093/nar/gkr035 (<https://doi.org/10.1093%2Fnar%2Fgkr035>). PMC 3082908 (<http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3082908>). PMID 21297115 (<https://pubmed.ncbi.nlm.nih.gov/21297115>).

External links

- [deltaMasses \(https://web.archive.org/web/20160418034750/http://www.detectorvision.com/deltaMasses/\)](https://web.archive.org/web/20160418034750/http://www.detectorvision.com/deltaMasses/) Detection of Methylations after Mass Spectrometry
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